

Gene expression data from nitrate-limited chemostat cultures of the diatom *Thalassiosira pseudonana* subjected to varying CO₂ concentrations

Website: <https://www.bco-dmo.org/dataset/546449>

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Project

» [A systems biology approach of diatom response to ocean acidification and climate change](#) (diatom response to OA and CC)

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Dataset Description

Gene expression data from nitrate-limited chemostat cultures of the diatom *Thalassiosira pseudonana* subjected to CO₂ concentrations ranging from 300-800 ppm.

Access to the dataset is restricted until the manuscript publication of our manuscript (currently under review for Nature Climate Change). Upon acceptance of the manuscript, data will be submitted to NCBI and the accession numbers will be provided here.

Methods & Sampling

For a full description of chemostat culturing methods see Hennon et al. (2014). Briefly, cells in chemostats were acclimated to 10 μ M nitrate limitation at 70% of max growth rate for >10 days (equivalent to >15 generations) at 300, 475 or 800 μ atm CO₂, verified by pH and DIC measurements. After steady-state acclimation, $\sim 1 \times 10^8$ cells were harvested on 0.2 micron polycarbonate filters by gentle vacuum filtration and flash frozen. Transition samples were collected daily as CO₂ levels were increased from 300-800 ppm over four consecutive days (equivalent to 6 generations) after pre-acclimation to 300 ppm CO₂ and nitrate limitation. $\sim 1.5 \times 10^7$ cells were harvested from each day of the transition experiment on 0.2 micron polycarbonate filters by gentle vacuum filtration and flash frozen. Each sample condition was reproduced with four biological replicates. A total of 28 barcoded transcriptome libraries were prepared; 12 libraries (three treatments x four biological replicates) from steady-state culture experiments and 16 libraries (four days x four biological replicates) from transition experiments. RNA was extracted from filtered cells using the ToTALLY RNA kit. Messenger RNA was selectively amplified using MessageAmp II aRNA Amplification kit creating the antisense aRNA complement to mRNA from the sample. SOLiD barcoded libraries were prepared from aRNA as described in SOLiD Total RNA-seq kit and sequenced on a SOLiD 5500XL sequencer in two runs: one containing steady-state barcoded libraries and one containing transition barcoded libraries.

Reference:

Hennon, G. M. M., Quay, P., Morales, R. L., Swanson, L. M., & Armbrust, E. V. 2014. Acclimation conditions modify physiological response of the diatom *Thalassiosira pseudonana* to elevated CO₂ concentrations in a nitrate-limited chemostat. *Journal of Phycology*, 253, 243–253. doi:[10.1111/jpy.12156](https://doi.org/10.1111/jpy.12156).

Data Processing Description

Reads were quality controlled (using a cut-off of $p = 0.99$ and minimum length = 30), trimmed, and aligned to *T. pseudonana* gene models using the Burrows-Wheeler Alignment tool and the SEAS_tAR tool (<https://github.com/armbrustlab/SEASAR>). The aligned reads were counted for each gene model (Joint Genome Institute: thaps3 extended models) using the SEAS_tAR tool.

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Parameters

Parameters for this dataset have not yet been identified

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Project Information

A systems biology approach of diatom response to ocean acidification and climate change (diatom response to OA and CC)

Description from the NSF award abstract:

Advances in systems biology enable new approaches to understand the carbon cycle and carbon sequestration research across different scales of organization, linking single cells to ecosystems, with broad impacts. Systems biology is a robust, holistic, hypothesis-driven, quantitative, integrative and iterative discipline that enables comprehensive understanding of model organisms by utilizing genomic and computational tools that provide the power for linking gene expression, phenotype, and the environment. In a systems approach, cells are studied as an integrated whole to explain the overall response and dynamic change in the full spectrum of molecules (DNA, RNA proteins and metabolites), and their relationships (biological networks). The publication of the genome of *T. pseudonana* allows a universal analysis and understanding of the regulation of primary production of diatoms in response to ocean acidification and climate change. Global understanding of the mechanisms of regulation of carbon fixation by diatoms is now possible. This project will focus on characterizing - at molecular and cellular levels using a systems approach - the response of diatoms to ocean acidification and climate change, essential to understanding the future of the ocean's "biological pump". The broader goal of this project is to understand the contribution by diatoms to carbon cycling at a biogeochemical level. This project will generate a model of the global expression of all genes in the diatom *T. pseudonana* and will enable us to anticipate how higher CO₂ and temperatures, lower pH will affect the ability of diatoms to sequester carbon in the oceans.

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Funding

Funding Source	Award
NSF Emerging Frontiers Division (NSF EF)	EF-1316206
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