

Weekly surface water samples from Newport Pier, CA collected between 11 January 2012 and 3 November 2023

Website: <https://www.bco-dmo.org/dataset/564351>

Data Type: Other Field Results

Version: 3

Version Date: 2025-03-05

Project

» [Biological Controls on the Ocean C:N:P ratios](#) (Biological C:N:P ratios)

Programs

» [Dimensions of Biodiversity](#) (Dimensions of Biodiversity)

» [Ocean Carbon and Biogeochemistry](#) (OCB)

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Abstract

Weekly surface water samples from Newport Pier, CA, were collected between 11 January 2012 and 3 November 2023. This project is a continuation of a time series with weekly surface water sampling from the Microbes in the Coastal Region of Orange County (MICRO) time series station at Newport Pier (33°36.37'N, 117°55.87'W). We measured dissolved nitrate and phosphate in the water, as well as particulate organic carbon, nitrogen and phosphorus. These data were published in Fagan et al. (2019), Larkin et al. (2020), and a pending publication, Larkin et al. (in press).

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Coverage

Location: Newport Pier, CA

Spatial Extent: Lat:33.6062 Lon:-117.9312

Temporal Extent: 2012-01-11 - 2023-11-03

Dataset Description

Associated sequence data are available at The National Center for Biotechnology Information (NCBI) under BioProject PRJNA624320 "Microbes In the Coastal Region of Orange County (MICRO) time series". This dataset includes NCBI accession numbers in the column "NCBI_BioSample". This BioSample accession information and related SRA data are accessible from the BioProject page <https://www.ncbi.nlm.nih.gov/bioproject/PRJNA624320/>.

Additional metagenomic data across the entire time series is available from the DOE Joint Genome Institute under "Detecting nutrient limitation and coastal biogeochemical responses to El Nino using microbial eco-genomic biomarkers" (award DOI: [10.46936/10.25585/60001365](https://doi.org/10.46936/10.25585/60001365)) accessible from page <https://genome.jgi.doe.gov/portal/Detnutbiomarkers/Detnutbiomarkers.info.html>.

Methods & Sampling

Sampling and analytical procedures:

Surface water from the "Microbes in the Coastal Region of Orange County" (MICRO) time series station at Newport Pier (33 36.37' N, -117 55.87' W) was collected weekly, in the morning, from 11 Jan 2012 to 26 December 2018. One liter polycarbonate bottles rinsed three times prior to sampling were filled for quantification of particulate organic matter and nutrient concentrations (three samples each from two bottles for a total of six replicates). Temperature, salinity, and chlorophyll a were continuously monitored using an automated shore station mounted next to the sampling site (www.sccoos.org).

Triplicate 300 ml samples for POC/PON or POP from each bottle were filtered within an hour of collection through pre-combusted (500 °C, 5 h) 25 mm GF/F filters (Whatman, MA). Each filter was rinsed with Milli-Q water before being fitted in order to remove potential P residues. The filtrate from the initial filtration was collected and used for macronutrient quantification. The filtrate was filtered through a 0.2 µm syringe filter into a 50 ml tube. Triplicates were collected for both macronutrient and stored at -20 °C.

Nitrate and phosphate samples were collected in prewashed 50 mL Falcon tubes and filtered through a 0.2 µm syringe filter and stored at -20 °C until further analysis. Soluble reactive phosphorus (SRP) concentrations were determined using the magnesium-induced co-precipitation (MAGIC) protocol and calculated against a potassium monobasic phosphate standard. (Karl and Tien, 1992; Lomas et al., 2010). Nitrate samples were treated with a solution of ethylenediaminetetraacetate and passed through a column of copperized cadmium fillings (Knap et al., 1993). Measurements were conducted using the same standards and protocols throughout the time series.

After thawing, POC/PON filters were allowed to dry overnight at 65 °C before being packed into a 30 mm tin capsule (CE Elantech, Lakewood, New Jersey). Samples were then analyzed for C and N content on the same FlashEA 1112 nitrogen and carbon analyzer (Thermo Scientific, Waltham, Massachusetts), following the Sharp (1974) protocol. POC and PON concentrations were calibrated using known quantities of atropine.

For published methodologies, see Allison et al. (2012), Fagan et al. (2019) and Martiny et al., (2015).

Data Processing Description

Issue report:

SCCOOS salinity meter down between March - May 2016.

Picoplankton columns are included in the dataset to maintain consistency with previously submitted data, but flow cytometry counts were not performed by our laboratory during the 2015-2018 time period.

BCO-DMO Processing Description

Version 3 (2025-03-05):

[data from 2018 to 2023 appended to the dataset, some parameters were removed and some have new

names, but the methods for the dataset are the same as prior versions]:

- * submitted file "NewportPier_MicrobeEnviro_2012-2023.csv" was imported into the BCO-DMO data system for this dataset. Values "NaN" and "N/A" were imported as missing data values. Submitters confirmed they indicate the same thing.

- * table filtered to remove columns that had "NaT" in the date column and all values except JGI_DOI blank.

Submitters clarified these should be removed and were remnant information.

- ** Missing data values are displayed differently based on the file format you download. They are blank in csv files, "NaN" in MatLab files, etc.

- * Column names adjusted to conform to BCO-DMO naming conventions designed to support broad re-use by a variety of research tools and scripting languages. [Only numbers, letters, and underscores. Can not start with a number]

- * Date converted to ISO 8601 format

- * All the numeric decimal columns were rounded to three decimal places except the picoeuk_ml which was rounded to 2 places as the rest of the *_ml parameters were already rounded to 2 places.

- * renamed column "NCBI_SRA_Accession" column name to "NCBI_BioSample" for clarity since it contains BioSample numbers. The SRA accessions will be accessible from the BioProject Page.

- * Provided submitter with the BCO-DMO revised file for review.

- * Submitter amended the file by providing additional JGI identifiers.

- * Saved this final file as "564351_v3_weekly-water-samples.csv".

Version 2 (2020-08-20): [data from 2015 to 2018 appended to version 1]

- * Extracted data submitted in Excel file BCODMO-Data_MICRO_NewportPier_CA-2015-2018.xlsx to csv with values as formatted in Excel (some numeric values had more precision than the set formatting).

- * Columns Month, Day and Year were removed from the 2015-2018 data. They were not in v1 and are redundant with information in Date column.

- * Added to data from version 1

- * blank values in this dataset are displayed as "nd" for "no data." nd is the default missing data identifier in the BCO-DMO system.

- * converted Date format to ISO 8601 format YYYY-MM-DD

Version 1 (2015-07-30):

- Generated from original file: "BCODMO_Newport.xlsx" contributed by Celine Mouginit

- Single column for Date inserted with combined Year,Month,Day formatted as YYYYMMDD

- Parameter names edited to conform to BCO-DMO naming convention found at Choosing Parameter Name

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Data Files

File
564351_v3_weekly-water-samples.csv (Comma Separated Values (.csv), 203.72 KB) MD5:eef59f0567a8f75dff7982a0e59decab
Primary data file for dataset ID 564351, version 3.

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Related Publications

Allison, S. D., Chao, Y., Farrara, J. D., Hatosy, S., & Martiny, A. C. (2012). Fine-Scale Temporal Variation in Marine Extracellular Enzymes of Coastal Southern California. *Frontiers in Microbiology*, 3.

doi:[10.3389/fmicb.2012.00301](https://doi.org/10.3389/fmicb.2012.00301)

Methods

Fagan, A. J., Moreno, A. R., & Martiny, A. C. (2019). Role of ENSO Conditions on Particulate Organic Matter Concentrations and Elemental Ratios in the Southern California Bight. *Frontiers in Marine Science*, 6.

doi:[10.3389/fmars.2019.00386](https://doi.org/10.3389/fmars.2019.00386)

Results

Karl, D. M., & Tien, G. (1992). MAGIC: A sensitive and precise method for measuring dissolved phosphorus in aquatic environments. *Limnology and Oceanography*, 37(1), 105–116. doi:[10.4319/lo.1992.37.1.0105](https://doi.org/10.4319/lo.1992.37.1.0105)
Methods

Knap, A., Michaels, R., Dow, R., Johnson, K., Gundersen, J., Sorensen, A., ... & Waterhouse, T. (1993). Bermuda Atlantic time-series study methods manual (Version 3). Bermuda Biological Station for Research, US JGOFS. https://www.researchgate.net/publication/245583966_Bermuda_Atlantic_Time-series_Study_Methods_Manual_Version_3
Methods

Larkin, A. A., Moreno, A. R., Fagan, A. J., Fowlds, A., Ruiz, A., & Martiny, A. C. (2020). Persistent El Niño driven shifts in marine cyanobacteria populations. *PLOS ONE*, 15(9), e0238405. <https://doi.org/10.1371/journal.pone.0238405>
Results

Larkin, A.A., Brock, M.L., Fagan, A.J., Moreno, A.R., Gerace, S.D., Lees, L.E., Suarez, S.A., Elie-Fadros, E.A., & Martiny, A. (in press). Climate-driven succession in marine microbiome biodiversity and biogeochemical function. *Nature Comm.*
Results

Lomas, M. W., Burke, A. L., Lomas, D. A., Bell, D. W., Shen, C., Dyhrman, S. T., & Ammerman, J. W. (2010). Sargasso Sea phosphorus biogeochemistry: an important role for dissolved organic phosphorus (DOP). *Biogeosciences*, 7(2), 695–710. doi:[10.5194/bg-7-695-2010](https://doi.org/10.5194/bg-7-695-2010)
Methods

Martiny, A. C., Talarmin, A., Mouginot, C., Lee, J. A., Huang, J. S., Gellene, A. G., & Caron, D. A. (2015). Biogeochemical interactions control a temporal succession in the elemental composition of marine communities. *Limnology and Oceanography*, 61(2), 531–542. doi:[10.1002/lno.10233](https://doi.org/10.1002/lno.10233)
Methods

SCCOOS (2024). A Science-Based Decision Support System. SOUTHERN CALIFORNIA COASTAL OCEAN OBSERVING SYSTEM. Retrieved June 24, 2024, from <https://sccoos.org/> www.sccoos.org
Methods

Sharp, J. H. (1974). Improved analysis for “particulate” organic carbon and nitrogen from seawater¹. *Limnology and Oceanography*, 19(6), 984–989. doi:[10.4319/lo.1974.19.6.0984](https://doi.org/10.4319/lo.1974.19.6.0984)
Methods

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Related Datasets

IsRelatedTo

Swartout, A., Martiny, A. (2021). Detecting nutrient limitation and coastal biogeochemical responses to El Nino using microbial eco-genomic biomarkers. Joint Genome Institute (JGI) Genome Portal version:8.18.177. The Regents of the University of California. Available from <https://genome.jgi.doe.gov/portal/Detnutbiomarkers/Detnutbiomarkers.info.html>

University of California - Irvine. Microbes In the Coastal Region of Orange County (MICRO) time series. (2020). In: NCBI:BioProject: PRJNA624320 [Internet]. Bethesda, MD: National Library of Medicine (US), National Center for Biotechnology Information; Available from: <http://www.ncbi.nlm.nih.gov/bioproject/PRJNA624320>.

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Parameters

Parameter	Description	Units
Date	Date (ISO 8601 format)	unitless

NO3_1	Nitrate replicate 1	micromolar (uM)
NO3_2	Nitrate replicate 2	micromolar (uM)
NO3_3	Nitrate replicate 3	micromolar (uM)
NO3_4	Nitrate replicate 4	micromolar (uM)
NO3_5	Nitrate replicate 5	micromolar (uM)
NO3_6	Nitrate replicate 6	micromolar (uM)
NO3_Avg	Average of nitrate replicates	micromolar (uM)
SRP_1	Soluble reactive phosphorus replicate 1	micromolar (uM)
SRP_2	Soluble reactive phosphorus replicate 2	micromolar (uM)
SRP_3	Soluble reactive phosphorus replicate 3	micromolar (uM)
SRP_4	Soluble reactive phosphorus replicate 4	micromolar (uM)
SRP_5	Soluble reactive phosphorus replicate 5	micromolar (uM)
SRP_6	Soluble reactive phosphorus replicate 6	micromolar (uM)
SRP_Avg	Average of soluble reactive phosphorus replicates	micromolar (uM)
POP_1	Particulate organic phosphorus replicate 1	micromolar (uM)
POP_2	Particulate organic phosphorus replicate 2	micromolar (uM)
POP_3	Particulate organic phosphorus replicate 3	micromolar (uM)
POP_4	Particulate organic phosphorus replicate 4	micromolar (uM)
POP_5	Particulate organic phosphorus replicate 5	micromolar (uM)

POP_6	Particulate organic phosphorus replicate 6	micromolar (uM)
POP_Avg	Average of particulate organic phosphorus replicates	micromolar (uM)
PON_1	Particulate organic nitrogen replicate 1	micromolar (uM)
PON_2	Particulate organic nitrogen replicate 2	micromolar (uM)
PON_3	Particulate organic nitrogen replicate 3	micromolar (uM)
PON_4	Particulate organic nitrogen replicate 4	micromolar (uM)
PON_5	Particulate organic nitrogen replicate 5	micromolar (uM)
PON_6	Particulate organic nitrogen replicate 6	micromolar (uM)
PON_Avg	Average of particulate organic nitrogen replicates	micromolar (uM)
POC_1	Particulate organic carbon replicate 1	micromolar (uM)
POC_2	Particulate organic carbon replicate 2	micromolar (uM)
POC_3	Particulate organic carbon replicate 3	micromolar (uM)
POC_4	Particulate organic carbon replicate 4	micromolar (uM)
POC_5	Particulate organic carbon replicate 5	micromolar (uM)
POC_6	Particulate organic carbon replicate 6	micromolar (uM)
POC_Avg	Average of particulate organic carbon replicates	micromolar (uM)
Temp	Sea temperature	Degrees Celsius (deg C)
Chlorophyll	Sea salinity	Practical Salinity Units (PSU)
Salinity	Chlorophyll-a concentration	micrograms per liter (ug/L)

picoeuk_ml	Picoeukaryote concentration	cells per milliliter (cells/ml)
PNEconc_ml	PNE, Pico/nano-eukaryotic phytoplankton concentration	cells per milliliter (cells/ml)
Proconc_ml	Prochlorococcus concentration	cells per milliliter (cells/ml)
Synconc_ml	Synechococcus concentration	cells per milliliter (cells/ml)
JGI_DOI	Joint Genome Institute (JGI) Award DOI for "Detecting nutrient limitation and coastal biogeochemical responses to El Nino using microbial eco-genomic biomarkers (Proposal ID: 507072)". Related sequence data in the JGI Genome Portal are accessible by clicking the "Site Award URL" on the JGI Award DOI page.	unitless
JGI_Project_Name	JGI sequencing project name	unitless
JGI_Sequencing_Project_ID	JGI sequencing project ID	unitless
JGI_Library_Name	JGI sequence library name	unitless
JGI_Project_ID	JGI analysis project ID	unitless
NCBI_BioSample	NCBI BioSample accession number	unitless

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Instruments

Dataset-specific Instrument Name	FlashEA 1112 Analyzer; Thermo Scientific, Waltham, Massachusetts
Generic Instrument Name	Elemental Analyzer
Generic Instrument Description	Instruments that quantify carbon, nitrogen and sometimes other elements by combusting the sample at very high temperature and assaying the resulting gaseous oxides. Usually used for samples including organic material.

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Deployments

MICRO_NewportPier_CA

Website	https://www.bco-dmo.org/deployment/632387
Platform	MICRO time series station at Newport Pier
Start Date	2012-01-11
End Date	2015-04-30

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Project Information

Biological Controls on the Ocean C:N:P ratios (Biological C:N:P ratios)

Coverage: western North Atlantic; 60N to 20N along 66W longitude; 20N to 15S in the tropical Pacific

One of the fundamental patterns of ocean biogeochemistry is the Redfield ratio, linking the stoichiometry of surface plankton with the chemistry of the deep ocean. There is no obvious mechanism for the globally consistent C:N:P ratio of 106:16:1 (Redfield ratio), especially as there is substantial elemental variation among plankton communities in different ocean regions. Thus, knowing how biodiversity regulates the elemental composition of the ocean is important for understanding the ocean and climate as a whole -- now and in the future.

The conceptual hypotheses for this study are as follows: 1. The C:N:P ratio of a cell is constrained by its broad taxonomic group, which determines, for example, whether it has an outer shell, its size, functional metabolism, membrane lipid composition. 2. Within a taxon, there is high genetic diversity. Some of this genetic diversity is potentially laterally transferred, or can be lost within taxa, and confers various functional abilities (organic phosphate assimilation, nitrate assimilation, photoheterotrophy, etc.). Functional diversity provides the cell with further flexibility, such as the ability to respond to varying nutrient supply rates/ratios, and affects a cell's C:N:P ratio within the range specified by the taxon. 3. Given these taxonomic and genetic constraints, a cell is physiologically plastic and modifies how it allocates cellular resources in response to nutrient supply rates/ratios in the environment. 4. The microbial diversity (taxonomic, genetic, and functional) of the surface ocean varies over time and space, driven by many factors in addition to nutrients. The sum of this mixture composes the ecosystem C:N:P, the ratio that Redfield described.

Based on this framework, the CoPIs will make field observations of taxon-specific stoichiometry and growth rates, genomic analyses, and conduct laboratory chemostat experiments to improve understanding of how ocean taxonomic, genetic, and functional biodiversity control the stoichiometry of the surface ocean plankton. Their analyses of these data would lead to a mechanistic understanding of variations in the Redfield ratio, both spatially and temporally.

This study will greatly expand knowledge of the genomic diversity among ocean microbes and how this diversity affects biogeochemistry. The stoichiometry of the ocean's microbes is a parameter that nearly every chemical or biological oceanographer uses, from converting measurements made in one element to another, to estimating regional and global nitrogen budgets. The research also has important implications for the global carbon budget and any changes that might result from climate change.

To understand mechanistically temporal and spatial variability of the plankton C:N:P ratio, biodiversity must be studied not only at the traditional taxonomic level, but at the genetic and functional levels which dictate organism response to their environment. Data will be integrated into a combined ocean ecological, evolutionary, and biogeochemical model, with flexible stoichiometry, including cellular biochemical allocations. Seeding a coupled physical-biological model of the oceans with multiple competing genotypes enables the exploration of ecological and evolutionary patterns of resource acquisition and C:N:P ratios. Developing a more mechanistic examination of the course of ecology and evolution, in which laboratory and field data define tradeoffs between different growth and nutrient acquisition strategies, would establish the framework of adaptive dynamics for determining "evolutionarily convergence". Finally, model outcomes will be evaluated against field data.

The field work planned for this project includes several cruises: BV46 (September/October 2011), BV48 (September 2012), a June 2013 cruise from Bermuda to the Labrador Sea, and a cruise from Hawaii to Tahiti

(May 2014). Additionally, samples will be acquired during cruises of opportunity.

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Program Information

Dimensions of Biodiversity (Dimensions of Biodiversity)

Website: http://www.nsf.gov/funding/pgm_summ.jsp?pims_id=503446

Coverage: global

(adapted from the NSF Synopsis of Program)

Dimensions of Biodiversity is a program solicitation from the NSF Directorate for Biological Sciences. FY 2010 was year one of the program. [\[MORE from NSF\]](#)

The NSF Dimensions of Biodiversity program seeks to characterize biodiversity on Earth by using integrative, innovative approaches to fill rapidly the most substantial gaps in our understanding. The program will take a broad view of biodiversity, and in its initial phase will focus on the integration of genetic, taxonomic, and functional dimensions of biodiversity. Project investigators are encouraged to integrate these three dimensions to understand the interactions and feedbacks among them. While this focus complements several core NSF programs, it differs by requiring that multiple dimensions of biodiversity be addressed simultaneously, to understand the roles of biodiversity in critical ecological and evolutionary processes.

Ocean Carbon and Biogeochemistry (OCB)

Website: <http://us-ocb.org/>

Coverage: Global

The Ocean Carbon and Biogeochemistry (OCB) program focuses on the ocean's role as a component of the global Earth system, bringing together research in geochemistry, ocean physics, and ecology that inform on and advance our understanding of ocean biogeochemistry. The overall program goals are to promote, plan, and coordinate collaborative, multidisciplinary research opportunities within the U.S. research community and with international partners. Important OCB-related activities currently include: the Ocean Carbon and Climate Change (OCCC) and the North American Carbon Program (NACP); U.S. contributions to IMBER, SOLAS, CARBOOCEAN; and numerous U.S. single-investigator and medium-size research projects funded by U.S. federal agencies including NASA, NOAA, and NSF.

The scientific mission of OCB is to study the evolving role of the ocean in the global carbon cycle, in the face of environmental variability and change through studies of marine biogeochemical cycles and associated ecosystems.

The overarching OCB science themes include improved understanding and prediction of: 1) oceanic uptake and release of atmospheric CO₂ and other greenhouse gases and 2) environmental sensitivities of biogeochemical cycles, marine ecosystems, and interactions between the two.

The OCB Research Priorities (updated January 2012) include: ocean acidification; terrestrial/coastal carbon fluxes and exchanges; climate sensitivities of and change in ecosystem structure and associated impacts on biogeochemical cycles; mesopelagic ecological and biogeochemical interactions; benthic-pelagic feedbacks on biogeochemical cycles; ocean carbon uptake and storage; and expanding low-oxygen conditions in the coastal and open oceans.

Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-1046001
NSF Division of Ocean Sciences (NSF OCE)	OCE-1046297