

# Sediment core sample metadata collected in the northern Gulf of Mexico, May 2017

**Website:** <https://www.bco-dmo.org/dataset/745789>

**Data Type:** Other Field Results

**Version:** 1

**Version Date:** 2018-10-15

## Project

» [Toward an Improved Understanding of Blue Carbon: The Role of Seagrasses in Sequestering CO<sub>2</sub>](#) (Seagrass Blue Carbon)

Contributors	Affiliation	Role
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## Abstract

This dataset includes metadata on sediment cores collected from the northern Gulf of Mexico in May 2017 - location, date and time of sample collection and processing, and which analyses were performed on each core.

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## Coverage

**Spatial Extent:** N:29.907 E:-84.456 S:29.853 W:-84.552

**Temporal Extent:** 2017-07-11 - 2017-07-20

## Dataset Description

This dataset includes metadata on sediment cores collected from the northern Gulf of Mexico in May 2017 - location, date and time of sample collection and processing, and which analyses were performed on each core.

## Methods & Sampling

Sediment cores were collected by divers, sealed in the field with rubber stoppers and returned to the lab for processing. Pore waters were collected by inserting rhizon samplers (Seeberg-Elverfeldt et al., 2005) through pre-drilled holes in the core tubes. Samples were collected in gas-tight glass syringes and filtered through 0.45 µm nylon filters into storage vials. Alkalinity samples were titrated within 12hr of collection; other samples were returned to the lab for analysis, using techniques routinely used in my lab.

Alkalinity and initial pH were determined by Gran Titration using a Metrohm automatic titrator (model 785 DMP

Titrimetric) combined with a Cole-Parmer pH electrode, calibrated using pH 4.00, 7.00 and 10.00 NIST-traceable buffers (Hu and Burdige, 2008). Sulfate was determined by ion chromatography and conductivity detection with a Thermo-Fisher Dionex ICS-5000 ion chromatograph, while DOC was determined by high-temperature combustion using a Shimadzu TOC-V total carbon analyzer (Burdige and Komada, 2011; Komada et al. 2016). Ammonium and DIC were determined by FIA analysis using a home-built system consisting of a Rainin Rabbit peristaltic pump and a Dionex CDM-II conductivity detector (Hall and Aller, 1992; Lustwerk and Burdige, 1995). Total dissolved sulfide was determined spectrophotometrically with an Ocean Optics USB400 UV-Vis spectrophotometer (Cline, 1969; Abdulla et al., in prep.); Total dissolved iron was also determined spectrophotometrically by the ferrozine method using the same spectrophotometer (Viollier et al., 2000).

Oxygen profiles in the cores were collected with polarographic microelectrodes (Luther et al., 2008) using a DLK 70 WebPstat electrochemical analyzer (AIS, Inc.) and a computer-controlled micro-profiler.

## Data Processing Description

BCO-DMO Processing Notes:

- added conventional header with dataset name, PI name, version date
- modified parameter names to conform with BCO-DMO naming conventions
- added columns for site, lat, and lon
- reformatted collection date and time from m/d/yyyy H:MM to YYYY-MM-DDTHH:MM:SS (ISO 8601:2004€ )
- replaced empty cell with - to distinguish them from x (all cells must be non-empty in our current system)
- changed Spidercrab Bay core id's from SC\* to SP\*

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## Data Files

File
<b>GoM_2017_core_meta.csv</b> (Comma Separated Values (.csv), 1.67 KB) MD5:1ffc3b7c70eed7686357ff77ff1ff5a7 Primary data file for dataset ID 745789

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## Related Publications

Abdulla, H. A., Burdige, D. J., & Komada, T. (2020). Abiotic formation of dissolved organic sulfur in anoxic sediments of Santa Barbara Basin. *Organic Geochemistry*, 139, 103879.

<https://doi.org/10.1016/j.orggeochem.2019.05.009>

*Methods*

Burdige, D. J., & Komada, T. (2011). Anaerobic oxidation of methane and the stoichiometry of remineralization processes in continental margin sediments. *Limnology and Oceanography*, 56(5), 1781–1796.

doi:[10.4319/lo.2011.56.5.1781](https://doi.org/10.4319/lo.2011.56.5.1781)

*Methods*

Cline, J. D. (1969). Spectrophotometric Determination of Hydrogen Sulfide in Natural Waters. *Limnology and Oceanography*, 14(3), 454–458. doi:[10.4319/lo.1969.14.3.0454](https://doi.org/10.4319/lo.1969.14.3.0454)

*Methods*

Hall, P. J., & Aller, R. C. (1992). Rapid, small-volume, flow injection analysis for total CO<sub>2</sub> and NH<sub>4</sub><sup>+</sup> in marine and freshwaters. *Limnology and Oceanography*, 37(5), 1113–1119. doi:[10.4319/lo.1992.37.5.1113](https://doi.org/10.4319/lo.1992.37.5.1113)

*Methods*

Hu, X., & Burdige, D. J. (2008). Shallow marine carbonate dissolution and early diagenesis—Implications from an incubation study. *Journal of Marine Research*, 66(4), 489–527. doi:[10.1357/002224008787157449](https://doi.org/10.1357/002224008787157449)

*Methods*

Komada, T., Burdige, D. J., Li, H.-L., Magen, C., Chanton, J. P., & Cada, A. K. (2016). Organic matter cycling

across the sulfate-methane transition zone of the Santa Barbara Basin, California Borderland. *Geochimica et Cosmochimica Acta*, 176, 259–278. doi:[10.1016/j.gca.2015.12.022](https://doi.org/10.1016/j.gca.2015.12.022)

*Methods*

Lustwerk, R. L., & Burdige, D. J. (1995). Elimination of dissolved sulfide interference in the flow injection determination of SCO<sub>2</sub>, by addition of molybdate. *Limnology and Oceanography*, 40(5), 1011–1012.

doi:[10.4319/lm.1995.40.5.1011](https://doi.org/10.4319/lm.1995.40.5.1011)

*Methods*

Luther, G. W., Glazer, B. T., Ma, S., Trouwborst, R. E., Moore, T. S., Metzger, E., ... Brendel, P. J. (2008). Use of voltammetric solid-state (micro)electrodes for studying biogeochemical processes: Laboratory measurements to real time measurements with an in situ electrochemical analyzer (ISEA). *Marine Chemistry*, 108(3-4), 221–235. doi:[10.1016/j.marchem.2007.03.002](https://doi.org/10.1016/j.marchem.2007.03.002)

*Methods*

Seeborg-Elverfeldt, J., Schlüter, M., Feseker, T., & Kölling, M. (2005). Rhizon sampling of porewaters near the sediment-water interface of aquatic systems. *Limnology and Oceanography: Methods*, 3(8), 361–371.

doi:[10.4319/lom.2005.3.361](https://doi.org/10.4319/lom.2005.3.361)

*Methods*

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## Parameters

Parameter	Description	Units
site	sample collection site identifier	unitless
lat	latitude; north is positive	decimal degrees
lon	longitude; east is positive	decimal degrees
core	core designator	unitless
date_processed	local date and time processed formatted as YYYY-MM-DDTHH:MM:SS (ISO 8601:2004E )	unitless
PW	pore water collection and analysis (alkalinity; pH; sulfate; DIC; DOC sulfide; ammonium; Fe) was performed	unitless
Ox	polarographic O <sub>2</sub> microsensor analysis was performed	unitless
phi	sediment porosity was measured	unitless
Pb210	<sup>210</sup> Pb analyses were performed	unitless
sol	solids were frozen from later analysis (TOC; Fe speciation; etc.)	unitless

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## Instruments

<b>Dataset-specific Instrument Name</b>	Dionex CDM-II conductivity detector
<b>Generic Instrument Name</b>	Conductivity Meter
<b>Dataset-specific Description</b>	Used to measure ammonium and dissolved inorganic carbon.
<b>Generic Instrument Description</b>	Conductivity Meter - An electrical conductivity meter (EC meter) measures the electrical conductivity in a solution. Commonly used in hydroponics, aquaculture and freshwater systems to monitor the amount of nutrients, salts or impurities in the water.

<b>Dataset-specific Instrument Name</b>	Thermo-Fisher Dionex ICS-5000 ion chromatograph
<b>Generic Instrument Name</b>	Ion Chromatograph
<b>Dataset-specific Description</b>	Used to measure sulfate.
<b>Generic Instrument Description</b>	Ion chromatography is a form of liquid chromatography that measures concentrations of ionic species by separating them based on their interaction with a resin. Ionic species separate differently depending on species type and size. Ion chromatographs are able to measure concentrations of major anions, such as fluoride, chloride, nitrate, nitrite, and sulfate, as well as major cations such as lithium, sodium, ammonium, potassium, calcium, and magnesium in the parts-per-billion (ppb) range. (from <a href="http://serc.carleton.edu/microbelife/research_methods/biogeochemical/ic....">http://serc.carleton.edu/microbelife/research_methods/biogeochemical/ic....</a> )

<b>Dataset-specific Instrument Name</b>	Ocean Optics USB400 UV-Vis spectrophotometer
<b>Generic Instrument Name</b>	Spectrophotometer
<b>Dataset-specific Description</b>	Used to measure total dissolved sulfide and total dissolved iron.
<b>Generic Instrument Description</b>	An instrument used to measure the relative absorption of electromagnetic radiation of different wavelengths in the near infra-red, visible and ultraviolet wavebands by samples.

<b>Dataset-specific Instrument Name</b>	Metrohm automatic titrator (model 785 DMP Titrino)
<b>Generic Instrument Name</b>	Titrator
<b>Dataset-specific Description</b>	Used to measure alkalinity and initial pH.
<b>Generic Instrument Description</b>	Titration is a process that incrementally adds quantified aliquots of a reagent to a sample until the end-point of a chemical reaction is reached.

<b>Dataset-specific Instrument Name</b>	Shimadzu TOC-V total carbon analyzer
<b>Generic Instrument Name</b>	Total Organic Carbon Analyzer
<b>Dataset-specific Description</b>	Used to measure dissolved organic carbon.
<b>Generic Instrument Description</b>	A unit that accurately determines the carbon concentrations of organic compounds typically by detecting and measuring its combustion product (CO <sub>2</sub> ). See description document at: <a href="http://bcodata.whoi.edu/LaurentianGreatLakes_Chemistry/bs116.pdf">http://bcodata.whoi.edu/LaurentianGreatLakes_Chemistry/bs116.pdf</a>

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## Project Information

### **Toward an Improved Understanding of Blue Carbon: The Role of Seagrasses in Sequestering CO<sub>2</sub> (Seagrass Blue Carbon)**

**Coverage:** Chesapeake Bay, Northern Gulf of Mexico, and Bahamas Banks

NSF abstract:

This research will develop a quantitative understanding of the factors controlling carbon cycling in seagrass meadows that will improve our ability to quantify their potential as blue carbon sinks and predict their future response to climate change, including sea level rise, ocean warming and ocean acidification. This project will advance a new generation of bio-optical-geochemical models and tools (ECHOES) that have the potential to be transform our ability to measure and predict carbon dynamics in shallow water systems.

This study will utilize cutting-edge methods for evaluating oxygen and carbon exchange (Eulerian and eddy covariance techniques) combined with biomass, sedimentary, and water column measurements to develop and test numerical models that can be scaled up to quantify the dynamics of carbon cycling and sequestration in seagrass meadows in temperate and tropical environments of the West Atlantic continental margin that encompass both siliciclastic and carbonate sediments. The comparative analysis across latitudinal and geochemical gradients will address the relative contributions of different species and geochemical processes to better constrain the role of seagrass carbon sequestration to global biogeochemical cycles. Specifically the research will quantify: (i) the relationship between C stocks and standing biomass for different species with different life histories and structural complexity, (ii) the influence of above- and below-ground metabolism on carbon exchange, and (iii) the influence of sediment type (siliciclastic vs. carbonate) on Blue Carbon storage. Seagrass biomass, growth rates, carbon content and isotope composition (above- and below-ground), organic carbon deposition and export will be measured. Sedimentation rates and isotopic composition of PIC, POC, and iron sulfide precipitates, as well as porewater concentrations of dissolved sulfide, CO<sub>2</sub>, alkalinity and salinity will be determined in order to develop a bio-optical-geochemical model that will predict the impact of seagrass metabolism on sediment geochemical processes that control carbon cycling in shallow waters. Model predictions will be validated against direct measurements of DIC and O<sub>2</sub> exchange in seagrass meadows, enabling us to scale-up the density-dependent processes to predict the impacts of seagrass distribution and density on carbon cycling and sequestration across the submarine landscape.

Status, as of 09 June 2016: This project has been recommended for funding by NSF's Division of Ocean Sciences.

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## Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	<a href="#">OCE-1635403</a>
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1633951</a>

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