Series 3A: Multiple stressor experiments on T. pseudonana (CCMP1014) - cell size estimated by forward scatter from flow cytometry

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Project
» Collaborative Research: Effects of multiple stressors on Marine Phytoplankton (Stressors on Marine Phytoplankton)

Contributors	Affiliation	Role
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Abstract
The experiments were designed to test the combined effects of three CO2 concentrations, four temperatures, and three light intensities on growth of the diatom T. pseudonana CCMP1014 in a multifactorial design. This dataset contains measurements of cell size expressed as forward scatter as well as in equivalent spherical diameter (ESD) in microns.

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Coverage

Temporal Extent: 2018-07-01 - 2018-10-31

Dataset Description

The experiments in Series 3A were designed to test the combined effects of three CO2 concentrations, four temperatures, and three light intensities on growth and photophysiology of the diatom T. pseudonana CCMP1014 in a multifactorial design. This dataset contains measurements of cell sizes as particle forward scatter from flow cytometry (and calculations of cell size - Equivalent Spherical Diameter, ESD).

Methods & Sampling

Three CO2 concentrations were tested: 410 ppm, 750 ppm, and 1000 ppm respectively. For each CO2 concentration, four temperatures were tested: 15 degrees-C, 20 degrees-C, 25 degrees-C, and 30 degrees-C. Within each temperature, three light levels were tested: a sub-optimum light (SOL) intensity of 60 umol photons · m-2 · s-1, an optimum light (OL) intensity of 400 umol photons · m-2 · s-1 and an extreme light (EL) intensity of 800 umol photons · m-2 · s-1. All lights were set at a 12 h day: 12 h dark cycle. For logistical reasons, experiments were partially conducted in series, with all light treatments at two temperatures (either 15 degrees-C and 25 degrees-C or 20 degrees-C and 30 degrees-C) running simultaneously. This was repeated for each CO2 concentration.

Experiments were conducted in Multicultivator MC-1000 OD units (Photon Systems Instruments, Drasov, Czech Republic). Each unit consists of eight 85 ml test-tubes immersed in a thermostated water bath, each independently illuminated by an array of cool white LEDs set at specific intensity and timing. A 0.2um filtered CO2-air mix (Praxair Distribution Inc.) was bubbled through sterile artificial seawater, and the humidified gas mix was supplied to each tube via gentle sparging through a 2um stainless steel diffuser. Flow rates were gradually increased over the course of the incubation to compensate for the DIC uptake of actively growing cells, and ranged from <0.04 Liters per minute (LPM) at the start of the incubations to 0.08 LPM in each tube after 2 days. For each CO2 and temperature level, replication was achieved by incubating three tubes at sub-optimum light intensities, two tubes at optimum light intensity, and three tubes at extreme light intensities. Each experiment was split into two phases: An acclimation phase spanning 4 days, was used to acclimate cultures to their new environment. Pre-acclimated, exponentially-growing cultures were then inoculated into fresh media and incubated through a 3-day experimental phase during which assessments of growth, photophysiology, and nutrient cycling were carried out daily. All sampling started 5 hours into the daily light cycle to minimize the effects of diurnal cycles.

Experiments were conducted with artificial seawater (ASW) prepared using previously described methods (Kester et. al 1967), and enriched with nitrate (NO3), phosphate (PO4), silicic acid (Si[OH]4), at levels ensuring that the cultures would remain nutrient-replete over the course of the experiment. Trace metals and vitamins were added as in f/2 (Guillard 1975). The expected DIC concentration and pH of the growth media was determined for the different pCO2 and temperatures using the CO2SYS calculator (Pierrot et al. 2006), with constants from Mehrbach et al. (1973, refit by Dickson & Millero 1987), and inputs of temperature, salinity, total alkalinity (2376.5 umol · kg-1), pCO2, phosphate, and silicic acid. DIC levels in ASW at the start of each phase of the experiments were manipulated by the addition of NaHCO3, and was then maintained by bubbling a CO2-Air mix through the cultures over the course of the experiments. The pH of the growth media was measured spectrophometrically using the m-cresol purple method (Dickson 1993), and adjusted using 0.1N HCl or 0.1M NaOH. The media was distributed into 75 ml aliquots and each aliquot was inoculated with 5 ml of the T. pseudonana CCMP 1014 (TP1014) stock culture at the start of the experiments.

Flow cytometry:
Samples were fixed in Hexamethylenetetramine-buffered formaldehyde (final concentration 1% v/v) and stored at 4 degrees C in the dark for a maximum of 4 days. Cell counts were confirmed to be unaffected over storage for up to a week. Samples were analyzed on a Guava easyCyte HT Benchtop Flow Cytometer (Millipore-Sigma, USA). All data acquisitions were done with logarithmic signal amplification. Cytometer sample flow rates were kept low (0.24 uL · s-1) to accommodate high cell concentrations. Diatoms were identified based on size and chlorophyll autofluorescence using the forward scatter channel (FSC) and Red-FL (695/50 nm) channel respectively. Growth rates were derived by fitting an exponential curve to cell concentrations vs. time for a 48-hour period during which cells exhibited exponential growth in the experimental phase. Growth rates in treatments where cells did not grow, or declined in abundance were listed as 0. Particle sizes (equivalent spherical diameter in µm, ESD) were derived from FSC using size-calibration beads of known diameters ranging from 2 µm to 10 um (Particle Size standard kit, Spherotech Inc.).

Data Processing Description

BCO-DMO Processing Notes:
- added conventional header with dataset name, PI name, version date
- modified parameter names to conform with BCO-DMO naming conventions
- changed "- NA -" to "NA"

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Data Files

File
3A_cellsize_expt.csv (Comma Separated Values (.csv), 5.45 KB) MD5:f99d60cfb631a12531e8ec72252820c8 Primary data file for dataset ID 771448

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Related Publications

Dickson, A. G. (1993). The measurement of sea water pH. Marine Chemistry, 44(2-4), 131–142. doi:10.1016/0304-4203(93)90198-w https://doi.org/10.1016/0304-4203(93)90198-W
Methods

Dickson, A. G., & Millero, F. J. (1987). A comparison of the equilibrium constants for the dissociation of carbonic acid in seawater media. Deep Sea Research Part A. Oceanographic Research Papers, 34(10), 1733–1743. doi:10.1016/0198-0149(87)90021-5
Methods

Dickson, A.G., Sabine, C.L. and Christian, J.R. (Eds.) 2007. Guide to best practices for ocean CO2 measurements. PICES Special Publication 3, 191 pp. ISBN: 1-897176-07-4. URL: https://www.nodc.noaa.gov/ocads/oceans/Handbook_2007.html https://hdl.handle.net/11329/249
Methods

Guillard, R. R. L. (1975). Culture of Phytoplankton for Feeding Marine Invertebrates. Culture of Marine Invertebrate Animals, 29–60. doi:10.1007/978-1-4615-8714-9_3
Methods

Kester, D. R., Duedall, I. W., Connors, D. N., & Pytkowicz, R. M. (1967). Preparation of Artificial Seawater 1. Limnology and Oceanography, 12(1), 176–179. doi:10.4319/lo.1967.12.1.0176
Methods

Mehrbach, C., Culberson, C. H., Hawley, J. E., & Pytkowicx, R. M. (1973). Measurement of the apparent dissociation constants of carbonic acid in seawater at atmospheric pressure. Limnology and Oceanography, 18(6), 897–907. doi:10.4319/lo.1973.18.6.0897
Methods

Pierrot, D. E. Lewis,and D. W. R. Wallace. 2006. MS Excel Program Developed for CO2 System Calculations. ORNL/CDIAC-105a. Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, U.S. Department of Energy, Oak Ridge, Tennessee. doi: 10.3334/CDIAC/otg.CO2SYS_XLS_CDIAC105a.
Methods

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Parameters

Parameter	Description	Units
Phase	Indicates whether the sample was collected during the acclimation phase or the experiment phase of the experiment.	unitless
CO2	Indicates the concentration of CO2 in the CO2-Air mix that was bubbled through the samples over the course of the experiment	parts per million (ppm)
Temperature	Indicates the temperature at which the samples were incubated.	degrees Celsius
Day	Indicates the timepoint (day) of sampling. D0 = day 0; D1 = day 1; etc.	unitless

Replicate	Indicates replication within a treatment. "NA" indicates "not applicable"	unitless
SOL_A_FSC	Forward scatter in replicate A incubated at sub optimum light (SOL)	FSC relative units
SOL_B_FSC	Forward scatter in replicate B incubated at sub optimum light (SOL)	FSC relative units
SOL_C_FSC	Forward scatter in replicate C incubated at sub optimum light (SOL)	FSC relative units
OL_A_FSC	Forward scatter in replicate A incubated at optimum light (OL)	FSC relative units
OL_B_FSC	Forward scatter in replicate B incubated at optimum light (OL)	FSC relative units
EL_A_FSC	Forward scatter in replicate A incubated at extreme light (EL)	FSC relative units
EL_B_FSC	Forward scatter in replicate B incubated at extreme light (EL)	FSC relative units
EL_C_FSC	Forward scatter in replicate C incubated at extreme light (EL)	FSC relative units
SOL_A_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate A incubated at sub optimum light (SOL)	micrometers
SOL_B_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate B incubated at sub optimum light (SOL)	micrometers
SOL_C_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate C incubated at sub optimum light (SOL)	micrometers
OL_A_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate A incubated at optimum light (OL)	micrometers
OL_B_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate B incubated at optimum light (OL)	micrometers
EL_A_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate A incubated at elevated light (EL)	micrometers
EL_B_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate B incubated at elevated light (EL)	micrometers
EL_C_ESD	Estimated Spherical Diameter (ESD) of cells calculated from forward scatter; replicate C incubated at elevated light (EL)	micrometers

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Instruments

Dataset-specific Instrument Name	Multicultivator MC-1000 OD (Qubit Systems)
Generic Instrument Name	Cell Cultivator
Dataset-specific Description	Used for incubation of TP1014 cultures.
Generic Instrument Description	An instrument used for the purpose of culturing small cells such as algae or bacteria. May provide temperature and light control and bubbled gas introduction.

Dataset-specific Instrument Name	Guava easyCyte HT Benchtop Flow Cytometer (Millipore-Sigma, USA)
Generic Instrument Name	Flow Cytometer
Dataset-specific Description	Used to measure abundance and forward scatter (proxy for cell size).
Generic Instrument Description	Flow cytometers (FC or FCM) are automated instruments that quantitate properties of single cells, one cell at a time. They can measure cell size, cell granularity, the amounts of cell components such as total DNA, newly synthesized DNA, gene expression as the amount messenger RNA for a particular gene, amounts of specific surface receptors, amounts of intracellular proteins, or transient signalling events in living cells. Description from: http://www.bio.umass.edu/micro/immunology/facs542/facswhat.htm

Dataset-specific Instrument Name	Genesys 10SVIS spectrophotometer
Generic Instrument Name	Spectrophotometer
Dataset-specific Description	Used to measure pH.
Generic Instrument Description	An instrument used to measure the relative absorption of electromagnetic radiation of different wavelengths in the near infra-red, visible and ultraviolet wavebands by samples.

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Project Information

Collaborative Research: Effects of multiple stressors on Marine Phytoplankton (Stressors on Marine Phytoplankton)

The overarching goal of this project is to develop a framework for understanding the response of

phytoplankton to multiple environmental stresses. Marine phytoplankton, which are tiny algae, produce as much oxygen as terrestrial plants and provide food, directly or indirectly, to all marine animals. Their productivity is thus important both for global elemental cycles of oxygen and carbon, as well as for the productivity of the ocean. Globally the productivity of marine phytoplankton appears to be changing, but while we have some understanding of the response of phytoplankton to shifts in one environmental parameter at a time, like temperature, there is very little knowledge of their response to simultaneous changes in several parameters. Increased atmospheric carbon dioxide concentrations result in both ocean acidification and increased surface water temperatures. The latter in turn leads to greater ocean stratification and associated changes in light exposure and nutrient availability for the plankton. Recently it has become apparent that the response of phytoplankton to simultaneous changes in these growth parameters is not additive. For example, the effect of ocean acidification may be severe at one temperature-light combination and negligible at another. The researchers of this project will carry out experiments that will provide a theoretical understanding of the relevant interactions so that the impact of climate change on marine phytoplankton can be predicted in an informed way. This project will engage high schools students through training of a teacher and the development of a teaching unit. Undergraduate and graduate students will work directly on the research. A cartoon journalist will create a cartoon story on the research results to translate the findings to a broader general public audience.

Each phytoplankton species has the capability to acclimatize to changes in temperature, light, pCO2, and nutrient availability - at least within a finite range. However, the response of phytoplankton to multiple simultaneous stressors is frequently complex, because the effects on physiological responses are interactive. To date, no datasets exist for even a single species that could fully test the assumptions and implications of existing models of phytoplankton acclimation to multiple environmental stressors. The investigators will combine modeling analysis with laboratory experiments to investigate the combined influences of changes in pCO2, temperature, light, and nitrate availability on phytoplankton growth using cultures of open ocean and coastal diatom strains (Thalassiosira pseudonana) and an open ocean cyanobacteria species (Synechococcus sp.). The planned experiments represent ideal case studies of the complex and interactive effects of environmental conditions on organisms, and results will provide the basis for predictive modeling of the response of phytoplankton taxa to multiple environmental stresses.

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Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-1538602

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