

Single colony metaproteomes of *Trichodesmium* from samples collected in North Atlantic surface waters during the R/V Atlantis cruise AT39-05 in March of 2018

Website: <https://www.bco-dmo.org/dataset/786694>

Data Type: Cruise Results

Version: 1

Version Date: 2020-01-10

Project

- » [Collaborative Research: Iron and phosphorus balanced limitation of nitrogen fixation in the oligotrophic ocean](#) (TriCoLim)
- » [New technology for high resolution analysis of proteins and other organic materials produced by marine microorganisms](#) (MM Proteins and Organics Tech)
- » [Marine Microbial Investigator Award: Investigator Mak Saito](#) (MM Saito)
- » [Collaborative Research: Evolutionary, biochemical and biogeochemical responses of marine cyanobacteria to warming and iron limitation interactions](#) (Cyanobacteria Warming Responses)

Program

- » [Marine Microbiology Initiative](#) (MMI)

Contributors	Affiliation	Role
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Abstract

Single colony metaproteomes of *Trichodesmium* from samples collected in North Atlantic surface waters during the R/V Atlantis cruise AT39-05 in March of 2018.

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Coverage

Spatial Extent: Lat:17.02 Lon:65.22

Temporal Extent: 2018-03-11

Dataset Description

Single colony metaproteomes of *Trichodesmium* from samples collected in North Atlantic surface waters during the R/V Atlantis cruise AT39-05 in March of 2018. These data were published in Held et al. (2020).

The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE [1] partner repository with the dataset identifier PXD016330 and 10.6019/PXD016330 but are not yet public.

* Project Name: *Trichodesmium* single colony metaproteomes field station Tricolim_19

* Project accession: PXD016330

* Project DOI: 10.6019/PXD016330

Methods & Sampling

A 200mm plankton net was deployed to ~20m depth, then recovered to just below surface, repeating five times. Trichodesmium colonies were hand-picked into 0.2mm filtered surface seawater, rinsed twice in 0.2mm filtered surface seawater, and decanted onto a 0.2-4mm Supor filter (indicated in sample provenance table, see dataset <https://www.bco-dmo.org/dataset/787093>).

Proteins were extracted and trypsin digested in-gel following Saito et al., 2014 (Science) with the following modifications: organic precipitation was skipped, benzonase nuclease treatment was performed instead to dissolve DNA/RNA material. Peptides were analyzed by LC-MS/MS in data discovery mode on a Thermo Orbitrap Fusion. Full details will be reported in Held et al. (in prep)

Location: North Atlantic surface waters 65.22 W 17.02 N

Data Processing Description

Thermo proteome discoverer 2.2 was used to search the data with the SEQUEST algorithm. Statistical validation was performed in Scaffold (Proteome Software) at the 1% protein and peptide false discovery rate (FDR) levels.

The sequence database was a trimmed set of Trichodesmium sequences which had been previously identified at this field station.

BCO-DMO Data Manager Processing Notes:

* originally submitted file TrichoSingleColony_dataset_TableS1.xlsx" converted to csv

* modified parameter names to conform with BCO-DMO naming conventions. "Product Name" changed to "Product_Name"

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Data Files

File	
Single colony metaproteomes of Trichodesmium	
filename: singlecolony.csv	(Comma Separated Values (.csv), 3.46 MB) MD5:05329123d1ee52317c4618b694eaf849
Single colony metaproteomes of Trichodesmium. See Parameters section for column names, descriptions, and units in this file.	

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Related Publications

Held, N. A., Webb, E. A., McIlvin, M. M., Hutchins, D. A., Cohen, N. R., Moran, D. M., ... Saito, M. A. (2020). Co-occurrence of Fe and P stress in natural populations of the marine diazotroph Trichodesmium. doi:[10.5194/bg-2019-493](https://doi.org/10.5194/bg-2019-493)
Results

ProteomeXchange dataset. (n.d.). doi:10.6019/pxd016330 <https://doi.org/10.6019/PXD016330>
Related Research

Saito, M. A., McIlvin, M. R., Moran, D. M., Goepfert, T. J., DiTullio, G. R., Post, A. F., & Lamborg, C. H. (2014). Multiple nutrient stresses at intersecting Pacific Ocean biomes detected by protein biomarkers. Science, 345(6201), 1173-1177.
<https://doi.org/10.1126/science.1256450>
Methods

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Related Datasets

IsSupplementedBy

Saito, M. (2020) **Trichodesmium sample provenance from samples collected in North Atlantic surface waters, station BATS, and station ALOHA between 2000 and 2018**. Biological and Chemical Oceanography Data Management Office (BCO-DMO). (Version 1) Version Date 2020-01-10 doi:10.26008/1912/bco-dmo.787093.1 [[view at BCO-DMO](#)]
Relationship Description: Sample provenance file, which includes sample locations, filter sizes.

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Parameters

Parameter	Description	Units
Protein_Accession	Protein accession, unique protein identifier.	unitless
Tricolim_14b	Relative protein abundance data for sample Tricolim_14b. Normalized MS1 Intensity.	unitless
Tricolim_15a	Relative protein abundance data for sample Tricolim_15a. Normalized MS1 Intensity.	unitless
Tricolim_16a	Relative protein abundance data for sample Tricolim_16a. Normalized MS1 Intensity.	unitless
Tricolim_16b	Relative protein abundance data for sample Tricolim_16b. Normalized MS1 Intensity.	unitless
Tricolim_15b	Relative protein abundance data for sample Tricolim_15b. Normalized MS1 Intensity.	unitless
Tricolim_15c	Relative protein abundance data for sample Tricolim_15c. Normalized MS1 Intensity.	unitless
Tricolim_15d	Relative protein abundance data for sample Tricolim_15d. Normalized MS1 Intensity.	unitless
Tricolim_18a	Relative protein abundance data for sample Tricolim_18a. Normalized MS1 Intensity.	unitless
Tricolim_18b	Relative protein abundance data for sample Tricolim_18b. Normalized MS1 Intensity.	unitless
Tricolim_18c	Relative protein abundance data for sample Tricolim_18c. Normalized MS1 Intensity.	unitless
JC150_02a	Relative protein abundance data for sample JC150_02a. Normalized MS1 Intensity.	unitless
JC150_02b	Relative protein abundance data for sample JC150_02b. Normalized MS1 Intensity.	unitless
Tricolim_03a	Relative protein abundance data for sample Tricolim_03a. Normalized MS1 Intensity.	unitless
Tricolim_03b	Relative protein abundance data for sample Tricolim_03b. Normalized MS1 Intensity.	unitless
Tricolim_13a	Relative protein abundance data for sample Tricolim_13a. Normalized MS1 Intensity.	unitless
Tricolim_19a	Relative protein abundance data for sample Tricolim_19a. Normalized MS1 Intensity.	unitless
Tricolim_19b	Relative protein abundance data for sample Tricolim_19b. Normalized MS1 Intensity.	unitless
Tricolim_19c	Relative protein abundance data for sample Tricolim_19c. Normalized MS1 Intensity.	unitless
JC150_03a	Relative protein abundance data for sample JC150_03a. Normalized MS1 Intensity.	unitless

JC150_03b	Relative protein abundance data for sample JC150_03b. Normalized MS1 Intensity.	unitless
JC150_03c	Relative protein abundance data for sample JC150_03c. Normalized MS1 Intensity.	unitless
JC150_04a	Relative protein abundance data for sample JC150_04a. Normalized MS1 Intensity.	unitless
JC150_05a	Relative protein abundance data for sample JC150_05a. Normalized MS1 Intensity.	unitless
JC150_05b	Relative protein abundance data for sample JC150_05b. Normalized MS1 Intensity.	unitless
JC150_05c	Relative protein abundance data for sample JC150_05c. Normalized MS1 Intensity.	unitless
JC150_06a	Relative protein abundance data for sample JC150_06a. Normalized MS1 Intensity.	unitless
JC150_07a	Relative protein abundance data for sample JC150_07a. Normalized MS1 Intensity.	unitless
JC150_07b	Relative protein abundance data for sample JC150_07b. Normalized MS1 Intensity.	unitless
JC150_07c	Relative protein abundance data for sample JC150_07c. Normalized MS1 Intensity.	unitless
BATS_01b	Relative protein abundance data for sample BATS_01b. Normalized MS1 Intensity.	unitless
BATS_01c	Relative protein abundance data for sample BATS_01c. Normalized MS1 Intensity.	unitless
JC150_01a	Relative protein abundance data for sample JC150_01a. Normalized MS1 Intensity.	unitless
JC150_01b	Relative protein abundance data for sample JC150_01b. Normalized MS1 Intensity.	unitless
JC150_01c	Relative protein abundance data for sample JC150_01c. Normalized MS1 Intensity.	unitless
HOT_01a	Relative protein abundance data for sample HOT_01a. Normalized MS1 Intensity.	unitless
HOT_01b	Relative protein abundance data for sample HOT_01b. Normalized MS1 Intensity.	unitless
BATS_01d	Relative protein abundance data for sample BATS_01d. Normalized MS1 Intensity.	unitless
Taxonomy	Taxonomic description. Taxonomy with semicolon as the delimiter (e.g. "Bacteria;Cyanobacteria;unclassified;Nostocales;Nostocaceae;Raphidiopsis;brookii;Raphidiopsis brookii D9")	unitless
Protein_Product_Name	Protein description	unitless
KO_Annotation	Kegg Ontology annotation	unitless

Pfam_Annotation	Pfam protein family annotation	unitless
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Instruments

Dataset-specific Instrument Name	Thermo Orbitrap Fusion mass spectrometer
Generic Instrument Name	Mass Spectrometer
Generic Instrument Description	General term for instruments used to measure the mass-to-charge ratio of ions; generally used to find the composition of a sample by generating a mass spectrum representing the masses of sample components.

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Deployments

AT39-05

Website	https://www.bco-dmo.org/deployment/765978
Platform	R/V Atlantis
Start Date	2018-02-11
End Date	2018-03-14
Description	For study of iron and phosphorus balanced limitation of nitrogen fixation in the oligotrophic ocean.

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Project Information

Collaborative Research: Iron and phosphorus balanced limitation of nitrogen fixation in the oligotrophic ocean (TriCoLim)

Coverage: Tropical Atlantic

NSF abstract:

Marine cyanobacteria are able to use or "fix" atmospheric nitrogen gas, and so supply much of the essential nutrient nitrogen that supports open ocean food chains. Oceanographers have usually thought that the growth of these nitrogen-fixing cyanobacteria is limited at any particular time and place by the supply of either iron, or of phosphorus. Preliminary experiments have shown, though, that these nitrogen fixers instead grow best when both iron and phosphorus are scarce at the same time. In this project, the researchers will use cellular indicators that are specific for iron and phosphorus limitation to determine how important this type of "balanced limitation" of nitrogen-fixing cyanobacteria is in controlling the productivity of ocean food chains in the tropical Atlantic Ocean. Two graduate students will be trained at the University of Southern California (USC) and Woods Hole Oceanographic Institution, as well as a postdoctoral researcher at USC. Educational outreach efforts will take place at a Los Angeles inner city high school with a student body that is over 98% Hispanic and African-American, and with underrepresented undergraduates in the USC Global Environmental Microbiology course. In addition, two Research Experiences for Undergraduates students will be supervised for summer research projects to help them learn about science career options.

The researchers will investigate the biological and biogeochemical consequences of this unique balanced iron/phosphorus-limited phenotype, using both laboratory and fieldwork approaches. During the first year of this project, the nitrogen-fixing cyanobacteria will be cultured under iron and/or phosphorus limitation, followed by application of proteomics and transcriptomics to identify genes that are potential diagnostic biomarkers for iron/phosphorus balanced limitation. Preliminary work has already identified one promising candidate biomarker in one cyanobacterium, an EzrA protein domain that appears to be associated with the cell size decreases seen specifically under balanced limitation, and the researchers have identified

numerous other potential candidates for similar biomarkers. During the second year, these new co-limitation biomarkers and others previously validated for iron limitation (IsiB) and phosphorus limitation (SphX) will be used to investigate balanced limitation during a research cruise transecting from relatively high-iron, low-phosphorus North Atlantic waters, to the relatively high-phosphorus, low-iron South Atlantic. This fieldwork component will survey nitrogen fixing cyanobacteria populations across this natural iron/phosphorus gradient for genetic, proteomic, and physiological indicators of balanced limitation, as well as testing their responses to iron and phosphorus manipulations in shipboard incubation experiments. The third year will be devoted to sample analysis, and publications exploring the responses of oceanic nitrogen fixers to simultaneous limitation by both iron and phosphorus.

New technology for high resolution analysis of proteins and other organic materials produced by marine microorganisms (MM Proteins and Organics Tech)

Website: <https://www.moore.org/grant-detail?grantId=GBMF3934>

In support of acquiring a high resolution mass spectrometer that incorporates the latest technologies for analyzing proteins and other organic materials.

Marine Microbial Investigator Award: Investigator Mak Saito (MM Saito)

In support of obtaining deeper knowledge of major biogeochemically relevant proteins to inform a mechanistic understanding of global marine biogeochemical cycles.

Collaborative Research: Evolutionary, biochemical and biogeochemical responses of marine cyanobacteria to warming and iron limitation interactions (Cyanobacteria Warming Responses)

NSF abstract:

The oceans absorb much of the heat generated by human activities, and this warming of the surface ocean has consequences for important groups of marine organisms. Marine cyanobacteria are one such key group of organisms, since they supply much of the essential carbon and nitrogen that supports nearly all the rest of the marine food web. Currently, the growth of cyanobacteria is mostly constrained by scarce supplies of the micronutrient element iron, but they are also very sensitive to the ongoing increases in seawater temperature. Preliminary results suggest that warming could partly mitigate the negative effects of iron limitation on marine cyanobacteria. This project examines in depth how these interactions between warming and iron limitation will affect the future ocean carbon and nitrogen cycles, using laboratory culture experiments showing how cyanobacteria respond to simultaneously changing temperature and iron supplies. Both short-term response studies and long-term evolutionary experiments testing for adaptation use a comprehensive set of molecular biology tools targeting genes to proteins. The final goal is to apply the results of these experiments to improve quantitative models predicting how the ocean's carbon and nitrogen cycles, biological productivity, and living resources will respond to a warming future climate. Two graduate students, a postdoc and 3-4 underrepresented undergraduate researchers are supported, and the investigators also mentor summer science interns from largely Hispanic local high schools.

The physiology, biochemistry and biogeography of nitrogen-fixing cyanobacteria and unicellular picocyanobacteria are strongly influenced by temperature, subjecting them to intense selective pressure as the modern ocean steadily warms up. These groups have likewise been rigorously selected under chronic iron (Fe) scarcity, and the availability of this crucial micronutrient is also changing with a shifting climate. This project examines short-term acclimation and long-term evolutionary responses of Fe-stressed marine cyanobacteria to a warmer environment. Preliminary data show that Iron Use Efficiencies (IUE, mols N fixed.hr⁻¹ mol cellular Fe⁻¹) of Fe-limited *Trichodesmium* increase 4 to 5-fold with a 5oC temperature increase, allowing the cells to much more efficiently leverage scarce available Fe supplies to grow and fix nitrogen. This means that warming can to a large degree mitigate the negative effects of Fe limitation on *Trichodesmium*, resulting in a modelled 22% increase in global nitrogen fixation by 2100 in a warmer climate. This project aims to uncover the cellular biochemical mechanisms involved in this Fe-limitation/thermal IUE effect in a four-year experimental evolution study of the diazotrophs *Trichodesmium* and *Crocospaera* and the picocyanobacteria *Synechococcus* and *Prochlorococcus*, under a multi-variate selection matrix of temperature and Fe availability. The objectives are to 1) Assess the long-term adaptive responses of fitness, IUE and physiology to Fe limitation and warming interactions in these four major cyanobacterial groups; 2) Determine the molecular and biochemical mechanisms behind the surprising Fe/warming interactive effect on IUE using genomics, transcriptomics and quantitative proteomics coupled with 'metalloproteomics' determinations of Fe content in critical proteins; 3) Compare and contrast acclimation and adaptation responses to Fe limitation and warming in key cyanobacteria taxa, and 4) Integrate results using a published biogeochemical modeling approach to assess global consequences for marine productivity and nitrogen fixation. This project offers a mechanistic and predictive understanding of adaptation to Fe and warming co-stressors in a rapidly changing future ocean environment for some of the most important photoautotrophic

functional groups in the ocean.

This award reflects NSF's statutory mission and has been deemed worthy of support through evaluation using the Foundation's intellectual merit and broader impacts review criteria.

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Program Information

Marine Microbiology Initiative (MMI)

Website: <https://www.moore.org/initiative-strategy-detail?initiativeld=marine-microbiology-initiative>

A Gordon and Betty Moore Foundation Program.

Forging a new paradigm in marine microbial ecology:

Microbes in the ocean produce half of the oxygen on the planet and remove vast amounts of carbon dioxide, a greenhouse gas, from the atmosphere. Yet, we have known surprisingly little about these microscopic organisms. As we discover answers to some long-standing puzzles about the roles that marine microorganisms play in supporting the ocean's food webs and driving global elemental cycles, we realized that we still need to learn much more about what these organisms do and how they do it—including how they evolved and contribute to our ocean's health and productivity.

The Marine Microbiology Initiative seeks to gain a comprehensive understanding of marine microbial communities, including their diversity, functions and behaviors; their ecological roles; and their origins and evolution. Our focus has been to enable researchers to uncover the principles that govern the interactions among microbes and that govern microbially mediated nutrient flow in the sea. To address these opportunities, we support leaders in the field through investigator awards, multidisciplinary team research projects, and efforts to create resources of broad use to the research community. We also support development of new instrumentation, tools, technologies and genetic approaches.

Through the efforts of many scientists from around the world, the initiative has been catalyzing new science through advances in methods and technology, and to reduce interdisciplinary barriers slowing progress. With our support, researchers are quantifying nutrient pools in the ocean, deciphering the genetic and biochemical bases of microbial metabolism, and understanding how microbes interact with one another. The initiative has five grant portfolios:

Individual investigator awards for current and emerging leaders in the field.

Multidisciplinary projects that support collaboration across disciplines.

New instrumentation, tools and technology that enable scientists to ask new questions in ways previously not possible.

Community resource efforts that fund the creation and sharing of data and the development of tools, methods and infrastructure of widespread utility.

Projects that advance genetic tools to enable development of experimental model systems in marine microbial ecology.

We also bring together scientists to discuss timely subjects and to facilitate scientific exchange.

Our path to marine microbial ecology was a confluence of new technology that could accelerate science and an opportunity to support a field that was not well funded relative to potential impact. Around the time we began this work in 2004, the life sciences were entering a new era of DNA sequencing and genomics, expanding possibilities for scientific research – including the nascent field of marine microbial ecology. Through conversations with pioneers inside and outside the field, an opportunity was identified: to apply these new sequencing tools to advance knowledge of marine microbial communities and reveal how they support and influence ocean systems.

After many years of success, we will wind down this effort and close the initiative in 2021. We will have invested more than \$250 million over 17 years to deepen understanding of the diversity, ecological activities and evolution of marine microbial communities. Thanks to the work of hundreds of scientists and others involved with the initiative, the goals have been achieved and the field has been profoundly enriched; it is now positioned to address new scientific questions using innovative technologies and methods.

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Funding

Funding Source	Award
Gordon and Betty Moore Foundation: Marine Microbiology Initiative (MMI)	GBMF3934
Gordon and Betty Moore Foundation: Marine Microbiology Initiative (MMI)	GBMF3782
NSF Division of Ocean Sciences (NSF OCE)	OCE-1657766
NSF Division of Ocean Sciences (NSF OCE)	OCE-1850719

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