

# The influence of reactive oxygen species on "respiration" isotope effect

**Website:** <https://www.bco-dmo.org/dataset/923859>

**Data Type:** experimental

**Version:** 1

**Version Date:** 2024-03-28

## Project

» [Clumped Oxygen Isotope Signature of Marine Dissolved Oxygen](#) (Microbial isotope effects)

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|---------------------------------|---|------------------------|
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## Abstract

The triple-oxygen isotope ( $^{17}\text{O}/^{16}\text{O}$ ,  $^{18}\text{O}/^{16}\text{O}$ ) measurement of oxygen-bearing species represents one of the most robust tools to directly trace oxygen cycling in the environment. One particularly consequential application of this isotope system is the analysis of dissolved oxygen ( $\text{O}_2$ ) in aquatic environments to determine gross oxygen production. This approach assumes that photosynthesis, microbial respiration, and gas exchange are the main drivers of dissolved  $\text{O}_2$  isotope compositions, and that each process is described by predictable, consistent triple-oxygen isotope effects. However, there currently exists a large disagreement in the literature on the triple-oxygen isotope effect of respiration, which carries major implications for global primary productivity estimates. Recent work has additionally highlighted the ubiquitous production of extracellular reactive oxygen species (ROS) such as superoxide and hydrogen peroxide by microorganisms; this flux maybe responsible for as much as 20% of net oxygen utilization in the ocean. To examine the influence of ROS-mediated  $\text{O}_2$  recycling on the oxygen utilization isotope effect, we measured the triple-oxygen isotope fractionations and mass laws of superoxide dismutase, catalase, and iron-mediated  $\text{H}_2\text{O}_2$  degradation. We incorporate these constraints into an oxygen isotope flux model to explore the influence of ROS-mediated oxygen cycling on "respiration" isotope effects in previous studies. We find that ROS-mediated oxygen cycling can reconcile the previously reported range of triple-oxygen isotope fractionation factors and that typical marine isotope effects are broadly consistent with independent estimates of superoxide-mediated oxygen loss. These data are described further in the related publication, Sutherland et al., 2022 (doi: 10.1016/j.gca.2022.02.033).

## Table of Contents

- [Dataset Description](#)
  - [Methods & Sampling](#)
  - [Data Processing Description](#)
  - [BCO-DMO Processing Description](#)
- [Data Files](#)
- [Related Publications](#)
- [Parameters](#)
- [Instruments](#)
- [Project Information](#)
- [Funding](#)

## Dataset Description

This dataset has been published "as is". Final review by the data submitter was not received after the data were imported into the BCO-DMO data system.

## Methods & Sampling

We have chosen a representative from each enzyme group to offer initial isotopic constraints on  $\text{O}_2$  evolved

from superoxide dismutase (SOD) and catalase. Specifically, we analyzed a representative CuZn SOD (Sigma S5395) and typical catalase (heme-binding, clade 3, small subunit, Sigma C1345). The same experiment apparatus was used to investigate the isotope effects of both enzymes. The reaction vessel consisted of a closed-system glass bulb with an inlet subject to continuous He flow (pre-scrubbed with 5A molecular sieve at liquid nitrogen temperature) through a glass frit, an outlet stream, and an injection port outfitted with a blue butyl rubber septum. The outlet stream was passed through multiple loops of stainless steel tubing held at liquid nitrogen temperature to trap any water vapor that escaped the reactor. KO<sub>2</sub> or H<sub>2</sub>O<sub>2</sub> were added for reactions with SOD and catalase, respectively. Effluent He carrier gas and product O<sub>2</sub> were then passed through a 5A molecular sieve trap to isolate O<sub>2</sub> gas evolved by the enzyme of interest.

In the case of catalase, 2 milligrams (mg) of enzyme was dissolved in approximately 50 milliliters (mL) of ultra-high purity (18 MX MilliQ) water and bubbled with He for a minimum of one hour to remove O<sub>2</sub>, at which point 30% H<sub>2</sub>O<sub>2</sub> was introduced to the solution through the injection port to an initial concentration of 3.9 millimolar (mM) in the reaction vessel. The reaction was allowed to proceed for a range of time intervals (a few minutes to several hours) to ensure adequate coverage of reaction progress for use with the Rayleigh equation. The oxygen isotope composition of the starting H<sub>2</sub>O<sub>2</sub> was determined using the same experimental setup (without catalase) by adding a slurry containing MilliQ water and KMnO<sub>4</sub> (amounting to a 10X excess of H<sub>2</sub>O<sub>2</sub>), which quantitatively oxidizes H<sub>2</sub>O<sub>2</sub> to O<sub>2</sub>. The average O<sub>2</sub> yield of the catalase and H<sub>2</sub>O<sub>2</sub> treatment was 48% (n = 3) that of the KMnO<sub>4</sub> treatment, consistent with the expected 2:1 stoichiometry of catalase.

SOD experiments were conducted with the same reaction vessel with some modifications. Given the rapid rate of reaction between superoxide and SOD, we instead characterized the oxygen isotope composition of the starting material (introduced as KO<sub>2</sub>) and the fully completed reaction to determine how oxygen isotopes are partitions between the oxidized and the reduced product. Methods for both measurements were modified from one previous study investigating the <sup>18</sup>O isotope effects of CuZn SOD. To measure the initial ROS oxygen isotope composition, several milligrams of KO<sub>2</sub> were introduced into the empty and dry reaction chamber, which was immediately flushed with He. Since water/moisture will cause KO<sub>2</sub> to spontaneously disproportionate, the powder was kept dry prior to starting the reaction. Separately, a He-sparged solution containing MilliQ water and an excess of K<sub>3</sub>[Fe(CN)<sub>6</sub>] was prepared and added to the chamber to quantitatively oxidize the starting material to O<sub>2</sub>. The SOD disproportionation reaction was performed similarly but using a solution of CuZn SOD, horse-radish peroxidase (HRP, to reduce product H<sub>2</sub>O<sub>2</sub> quantitatively to water), and K<sub>4</sub>[Fe(CN)<sub>6</sub>]•3H<sub>2</sub>O (reducing equivalents for HRP; Smirnov and Roth, 2006). To further survey ROS reactions in the environment and their impact on the oxygen isotope systematics of dissolved O<sub>2</sub>, we examined one non-enzymatic ROS decay pathway - Fe-mediated H<sub>2</sub>O<sub>2</sub> degradation. We used the methods outlined in Dole et al. (1952) with the same experimental apparatus described above to explore this Fe-catalyzed pathway. Reaction progress was monitored by comparing the amount of O<sub>2</sub> evolved from the reaction vessel relative to that from the KMnO<sub>4</sub> treatment.

### Instruments:

Following O<sub>2</sub> gas collection, the sample volume was sealed with gas-tight valves under He flow and immediately transferred to a gas purification manifold to be analyzed on a Thermo Scientific MAT 253 Plus isotope ratio mass spectrometer (IRMS) at Harvard University (Cowie and Johnston, 2016). Carrier He gas was pumped away from the still-frozen sample before thawing. Oxygen gas should be the only product gas evolved from these reactions; however, to ensure trace atmospheric contaminants were not present, each sample was passed through a 3 m gas chromatography (GC) column packed with 5A molecular sieve maintained at 30 °C with He carrier gas at 15 mL/min. Oxygen was collected from GC effluent on a U-trap containing 5A molecular sieve held at liquid nitrogen temperature. Since all sample O<sub>2</sub> was generated from KO<sub>2</sub> and H<sub>2</sub>O<sub>2</sub>, Ar - which can interfere with TOI analyses - was not present (Yeung et al., 2018; Ash et al., 2020). Finally, effluent O<sub>2</sub> gas was cryo-focused and allowed to thaw at room temperature for a minimum of 20 min to ensure no isotope fractionation before introduction into the IRMS for analysis. Each analysis comprises the average value of four acquisition blocks, each consisting of 20 cycles between the reference gas and sample gas (total counting time on sample gas was 400 s per acquisition). Measurements were typically run at 5000 mV signal on the m/z 32 Faraday cup (3 x 10<sup>8</sup> resistor). Total acquisition time for a single analysis is roughly two hours.

### Data Processing Description

All values of d<sup>17</sup>O and d<sup>18</sup>O were converted to a VSMOW2-SLAP2 scale using a two point calibration of O<sub>2</sub> liberated from VSMOW2 and SLAP2 standards using a CoF<sub>3</sub> reactor as previously described (Schoenemann et al., 2013; Barkan and Luz, 2005). All slopes (i.e., k values) and slope uncertainties were calculated as linear fits to plots of d<sup>17</sup>O vs. d<sup>18</sup>O using the polyfit function in MATLAB (linear least-squares). Typical reproducibility of Ar-free lab air is 0.005‰ for D<sup>17</sup>O and 0.02‰ for d<sup>18</sup>O. The oxygen isotope results presented in this

study are the mean and standard deviation when the number of samples receiving the exact same treatment is greater than one. In situations where replicates were not possible (e.g., measurements along a reaction progress array), uncertainty is assumed to be equal to the reproducibility of lab air reported above.

## BCO-DMO Processing Description

- Imported original file "Sutherland GCA table.xlsx" into the BCO-DMO system.
- Renamed fields to comply with BCO-DMO naming conventions.
- Saved the final file as "923859\_v1\_reactive\_o2\_species.csv".
- This dataset has been published "as is". Final review by the data submitter was not received after the data were imported into the BCO-DMO data system.

[ [table of contents](#) | [back to top](#) ]

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## Data Files

| File  |
|---|
| <b>923859_v1_reactive_o2_species.csv</b> (Comma Separated Values (.csv), 1.37 KB)<br>MD5:134c9c9c462bce7e62004084f89fb86d |
| Primary data file for dataset ID 923859, version 1  |

[ [table of contents](#) | [back to top](#) ]

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## Related Publications

Ash, J. L., Hu, H., & Yeung, L. Y. (2019). What Fractionates Oxygen Isotopes during Respiration? Insights from Multiple Isotopologue Measurements and Theory. *ACS Earth and Space Chemistry*, 4(1), 50–66.

<https://doi.org/10.1021/acsearthspacechem.9b00230>

*Methods*

Barkan, E., & Luz, B. (2005). High precision measurements of  $^{17}\text{O}/^{16}\text{O}$  and  $^{18}\text{O}/^{16}\text{O}$  ratios in  $\text{H}_2\text{O}$ . *Rapid Communications in Mass Spectrometry*, 19(24), 3737–3742. Portico. <https://doi.org/10.1002/rcm.2250>

*Methods*

Cowie, B. R., & Johnston, D. T. (2016). High-precision measurement and standard calibration of triple oxygen isotopic compositions ( $\delta^{18}\text{O}$ ,  $\Delta^{17}\text{O}$ ) of sulfate by F2 laser fluorination. *Chemical Geology*, 440, 50–59.

<https://doi.org/10.1016/j.chemgeo.2016.07.003>

*Methods*

Dole, M., Rudd, D. P., Muchow, G. R., & Comte, C. (1952). Isotopic Composition of Oxygen in the Catalytic Decomposition of Hydrogen Peroxide. *The Journal of Chemical Physics*, 20(6), 961–968.

<https://doi.org/10.1063/1.1700657>

*Methods*

Schoenemann, S. W., Schauer, A. J., & Steig, E. J. (2013). Measurement of SLAP2 and GISP  $\delta^{17}\text{O}$  and proposed VSMOW-SLAP normalization for  $\delta^{17}\text{O}$  and  $^{17}\text{O}$  excess. *Rapid Communications in Mass Spectrometry*, 27(5), 582–590. Portico. <https://doi.org/10.1002/rcm.6486>

*Methods*

Smirnov, V. V., & Roth, J. P. (2006). Mechanisms of Electron Transfer in Catalysis by Copper Zinc Superoxide Dismutase. *Journal of the American Chemical Society*, 128(51), 16424–16425.

<https://doi.org/10.1021/ja066369r>

*Methods*

Sutherland, K. M., Hemingway, J. D., & Johnston, D. T. (2022). The influence of reactive oxygen species on “respiration” isotope effects. *Geochimica et Cosmochimica Acta*, 324, 86–101.

<https://doi.org/10.1016/j.gca.2022.02.033>

*Results*

Yeung, L. Y., Hayles, J. A., Hu, H., Ash, J. L., & Sun, T. (2018). Scale distortion from pressure baselines as a

source of inaccuracy in triple-isotope measurements. Rapid Communications in Mass Spectrometry, 32(20), 1811–1821. doi:[10.1002/rcm.8247](https://doi.org/10.1002/rcm.8247)  
*Methods*

[ [table of contents](#) | [back to top](#) ]

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## Parameters

| Parameter         | Description                                 | Units    |
|-------------------|---|----------|
| Sample            | Experimental label                          | unitless |
| n                 | Number of replicates                        | unitless |
| d18O_smowslap     | The 18O/16O ratio on a SMOW scale           | permil   |
| D17O              | The capital delta value                     | permil   |
| d17O_smowslap_log | The 17O/16O ratio on a SMOW scale, log form | unitless |
| d18O_smowslap_log | The 18O/16O ratio on a SMOW scale, log form | unitless |

[ [table of contents](#) | [back to top](#) ]

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## Instruments

|   |   |
|---|---|
| <b>Dataset-specific Instrument Name</b> | Faraday cup   |
| <b>Generic Instrument Name</b>          | Faraday cup   |
| <b>Generic Instrument Description</b>   | A Faraday cup is a metal (conductive) cup designed to catch charged particles in a vacuum. The resulting current can be measured and used to determine the number of ions or electrons hitting the cup. |

|   |  |
|---|--|
| <b>Dataset-specific Instrument Name</b> | 3 m gas chromatography (GC) column   |
| <b>Generic Instrument Name</b>          | Gas Chromatograph  |
| <b>Generic Instrument Description</b>   | Instrument separating gases, volatile substances, or substances dissolved in a volatile solvent by transporting an inert gas through a column packed with a sorbent to a detector for assay. (from SeaDataNet, BODC) |

|   |  |
|---|--|
| <b>Dataset-specific Instrument Name</b> | Thermo Scientific MAT 253 Plus isotope ratio mass spectrometer (IRMS)  |
| <b>Generic Instrument Name</b>          | Isotope-ratio Mass Spectrometer  |
| <b>Generic Instrument Description</b>   | The Isotope-ratio Mass Spectrometer is a particular type of mass spectrometer used to measure the relative abundance of isotopes in a given sample (e.g. VG Prism II Isotope Ratio Mass-Spectrometer). |

[ [table of contents](#) | [back to top](#) ]

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## Project Information

### Clumped Oxygen Isotope Signature of Marine Dissolved Oxygen (Microbial isotope effects)

#### NSF Award Abstract:

The balance between photosynthetic production of oxygen and biological consumption of oxygen in the marine environment plays a critical role in regulating the composition of Earth's atmosphere and the long-term stability of Earth's climate. The ability to accurately measure the production and consumption of oxygen in the marine environment is central to building an informed understanding of the past, present, and future of oxygen and Earth's climate. Measurement of the abundance of different oxygen isotopes (i.e. oxygen-16, oxygen-17, and oxygen-18) in dissolved oxygen in seawater is a powerful analytical tool that can be used to determine the magnitude of photosynthesis and biological oxygen consumption. This is possible because biological reactions that produce or consume oxygen tend to preferentially utilize different isotopes of oxygen. This analytical tool has been used for two decades to investigate ocean primary productivity, however, there is still significant uncertainty in how biogeochemical processes such as respiration preferentially select and utilize different oxygen isotopes. To remedy this uncertainty, researchers at Harvard University will perform a lab-based calibration and sea-going field deployment of emerging oxygen isotope analyses that target molecules of oxygen that contain two rare oxygen isotopes inside of one molecule (a.k.a. "clumps", i.e. 17O18O and 18O18O). This work will quantify the clumped oxygen isotope signatures of several of the most consequential oxygen-involving reactions that occur in the marine ecosystem. These oxygen isotope signatures will be used to refine current methods and assumptions for the measurement of gross oxygen production in the global ocean. This research will also help train the next generation of Earth scientists through the mentorship of one graduate student and two undergraduate students. This project will also facilitate the participation of researchers in content creation for a national science competition for middle and high school students that reaches thousands of students annually.

The application of clumped O2 isotope measurements to dissolved oxygen in seawater is poised to give a greater mechanistic view of gross primary productivity and marine oxygen cycling than previously attainable. To realize the analytical potential of clumped oxygen isotope methods, these researchers will characterize the clumped oxygen isotope effects associated with enzyme-level reactions, whole organisms, and the marine water column. Enzyme-level studies will include isotope characterization of a terminal-O2 reductase and enzymes that metabolize reactive oxygen species such as superoxide and hydrogen peroxide. Organism studies will target common and numerically abundant phototrophs and heterotrophs to explore the expected breadth of isotope signatures and fractionation factors in the marine water column. Marine water column dissolved oxygen samples will be collected during one of multiple candidate cruises and analyzed using both traditional triple oxygen isotope techniques and newly developed clumped oxygen isotope methods. Lastly, they will employ the results of lab-based study of clumped O2 to build a model for the interpretation of environmental data with the aim of improving the accuracy and precision of field measurements of gross primary production.

This award reflects NSF's statutory mission and has been deemed worthy of support through evaluation using the Foundation's intellectual merit and broader impacts review criteria.

[ [table of contents](#) | [back to top](#) ]

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## Funding

| Funding Source   | Award                       |
|--|-----------------------------|
| <a href="#">NSF Division of Ocean Sciences (NSF OCE)</a> | <a href="#">OCE-2049298</a> |

[ [table of contents](#) | [back to top](#) ]